Cold Atmospheric Pressure Plasma Jet and Plasma Lamp Interaction with Plants: Electrostimulation, Reactive Oxygen and Nitrogen Species, and Side Effects

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Abstract

Cold atmospheric pressure plasma (CAPP) treatment is a highly effective method of protecting seeds, plants, flowers, and trees from diseases and infection and significantly increasing crop yields.

Here we found that cold atmospheric pressure He-plasma jet (CAPPJ) can also cause side effects and damage to plants if the plasma exposure time is too long. Reactive oxygen and nitrogen species (RONS), electromagnetic fields, and ultraviolet photons emitted by CAPPJ can cause both positive and negative effects on plants. CAPPJ can interact with biological tissue surfaces. The plasma lamp has no visible side effects on Aloe vera plants, cabbage, and tomatoes. A plasma lamp and a cold atmospheric pressure plasma He-jet cause strong electrical signaling in plants with a very high amplitude with frequencies equal to the frequency of plasma generation. The use of plasma lamps for electrostimulation of biological tissues can help to avoid side processes in biological tissues associated with the generation of RONS, UV photons, and direct interaction with cold plasma. CAPP technology can play an important role in agriculture, medicine, the food industry, chemistry, surface science, material science, and engineering applications without side effects if the plasma exposure is short enough.

Introduction

CAPP treatment is a highly effective method of protecting bio-tissue from diseases and infection.

CAPP accelerates the imbibition and germination of seeds, plant growth, and nutrient absorption, activates enzymatic and ion channel activities [1-7], as well as promotes a significant increase in crop yields by up to 23% [2]. CAPP and plasma lamps can induce electroporation, corrugation, and morphological changes in the surfaces of seeds and biological tissues [4-6], as well as affect ion transport and bioelectrochemical characteristics of plant tissue [6,8-10].

There are also a few publications about side effects such as genotoxic effects, oxidation, and peroxidation of bio-
tissue induced by direct treatment of bio-tissue with cold atmospheric pressure plasma [8,10-17].

A plasma lamp (also called a plasma ball or Tesla ball) is a clear borosilicate glass ball filled with a combination of noble gases at atmospheric pressure with an electrode in the center of the sphere. Plasma lamps were developed by Tesla [18] and their physical properties were investigated recently [19]. A plasma lamp was already used for the electrostimulation of seeds and plants [5,8,20]. Plasma lamps produce strong electromagnetic fields and oscillating visible light which can interact with a bio-tissue [19]. Plasma in plasma lamps is separated and covered by glass from a bio-tissue. It does not produce RONS outside of the glass.

The effect of electric fields on vegetation has been the
subject of research since the eighteenth century [21-30]. Seed treatment with high-frequency electromagnetic fields using a plasma lamp can accelerate seed absorption, germination, and root growth without visible side effects [5,8,20]. Generated by plasma lamps or cold atmospheric pressure He-plasma jet (CAPPJ), high-frequency electromagnetic fields and photons can penetrate seed coats and modify their surface properties [5,8,20]. Treatment with a plasma lamp is not as effective for a harvest as treatment with a CAPPJ but usually does not generate side effects. Plasma lamps can be used in plasma agriculture to accelerate the germination of seeds, the growth of plant seedlings, and the corrugation of the surfaces of biological tissues without the side effects of reactive oxygen and nitrogen species (RONS) generated by plasma jets [5,8,20].

In this study, we tried to study possible side effects and electrical signaling in Aloe vera L., Brassica oleracea L., and Lycopersicon esculentum Mill. Plants caused by a cold atmospheric pressure He-plasma jet and a plasma ball. These plants are model objects for the study of electrical signaling and memory in plants [26,27,31-33].

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**Materials and methods**

**Plants**

Forty Aloe vera L. plants were grown in clay pots with sterilized potting soil. Plants were exposed to a 12:12 hr light/dark photoperiod at 21 °C. Aloe vera L. plants had 20 - 35 cm leaves. The soil around the Aloe vera L. plants was treated with water every week. Aloe vera plants were obtained from Bioelectrochemistry LLC (Madison, Al., USA).

Seedlings of Bonnie Hybrid Bonnie’s Best Cabbage (Brassica oleracea L.) were purchased from Bonnie Plant Farm (Union Spring, Al., USA). The soil around the cabbage plants was treated with water 3 times a week.

Forty tomatoes (Lycopersicon esculentum Mill. cv Cosmonaut Volkov) plants were grown in plastic pots with sterilized potting soil in a plant growth chamber (Environmental Corporation, USA). The soil around the tomato plants was treated with water every week. All measurements were performed on 21-to 28-day-old tomato plants. The seeds were purchased from various sources in Ukraine and Russia.

Irradiance was 850 μmol - 1100 μmol photons m⁻²s⁻¹ PAR at plant level. All experiments were performed on healthy specimens. The relative humidity in the laboratory was kept at 45% - 50%.

**Chemicals and test strips**

Ozone test strips were purchased from Macherey-Nagel Company (Duren, Germany). These were used to determine the ozone concentration in the air near the surface of a plasma ball and under the plasma jet (Figure 2). Bottled ultra-high purity helium was purchased from Sexton Welding Supply (Huntsville, Al., USA).

**Shielded extracellular electrodes**

Teflon-coated silver wires (A-M Systems, Inc., Sequim, WA, USA) with a diameter of 0.2 mm were used for the preparation of non-polarizable Ag/AgCl electrodes. Identical shielded electrodes (Ag/AgCl or Cu) were used as working and reference electrodes for measurements of electrical potential differences in the plants.

**Data acquisition**

The experimental setup for tests using a plasma lamp is shown in Figure 1. High-speed data acquisition was performed using NI-PXI-1042Q microcomputers with simultaneous multifunction I/O plug-in data acquisition board NI-PXI-6151 (National Instruments, Austin, TX, USA) interfaced through a NI-SCB-68 shielded connector block to Ag/AgCl electrodes. Data acquisition board NI-PXI-6151 (National Instruments, Austin, TX, USA) had a maximum sampling rate of 4,000,000 samples/s. The data acquisition board was connected to shielded electrodes.

**Plasma ball and cold atmospheric pressure He-plasma jet**

Common commercial plasma Nebula Plasma Ball (Figure 1) was used for electrostimulation of plants. The electromagnetic interference was measured with a CalTest CT2982B 10 kV high voltage divider probe (CalTest electronics, Yorba Linda, CA, USA) connected to a LeCroy wave master LT322 oscilloscope (LeCroy, Chesnut Ridge, NY, USA).

The CAPPJ method was described earlier [6,7]. The system was operated with 8 kV pulse amplitude, 6 kHz pulse frequency, 1 μs pulse width, and a ~70 ns rise time and fall time. The entire system is placed in a metal enclosure to reduce electromagnetic interference.

**FTIR spectra**

FTIR spectra were recorded using a Thermo Scientific Nicolet ISS FT-IR spectrometer (ThermoFisher Scientific, Waltham, Massachusetts, USA). Reflectance spectra were recorded on a spectrophotometer ISR-2000 Plus with an integrating sphere (Shimadzu, Japan).

**Temperature control**

Digital laser temperature gun Etekcity laser grip 800 (Etekcity, Anaheim, CA, USA) was used to measure the temperature of the plasma jet, water, plants, and air. The temperature of the plasma jet, plants, and air during plasma treatment was 20 °C.
Cold Atmospheric Pressure Plasma Jet and Plasma Lamp Interaction with Plants: Electrostimulation, Reactive Oxygen and Nitrogen Species, and Side Effects

Statistics

All experimental results were reproduced at least 14 times using different plants of *Aloe vera* L., *Brassica oleracea* L. and *Lycopersicon esculentum* Mill. cv Cosmonaut Volkov.

Results

Plasma lamps interaction with plants: Electrostimulation

If a plant is placed on the outer surface of the ball or near the ball (Figure 1), the capacitive coupling can supply a high voltage load of up to several kV with a frequency of about 22 kHz. The electrical signal generated by the plasma ball is not limited to the glass sphere and propagates into the surrounding air in the form of electromagnetic interference (EMI). The amplitude of the electrical signal from the probe at a distance of 1 cm from the plasma ball was 628 V. UV-Vis radiation with a wavelength of more than 380 nm can penetrate through the glass wall of the plasma ball. While the plasma inside the ball is surrounded by the glass wall and does not produce a significant amount of RONS outside the lamp, high-frequency electromagnetic radiation propagates outside. The measurement of the ozone level on the surface of our plasma balls conducted with commercial ozone test strips exposed for 10 min did not show the formation of ozone in the air near the glass surface (Figure 2A,B). The ozone test strips, when placed under a cold He-plasma jet at a distance of 1 cm for 10 min, showed the production of ozone by the He-plasma jet at a concentration of more than 200 μg/m³ (Figure 2C).

A monocot *Aloe vera* (L.) belongs to the Asphodelaceae (Liliaceae) family with crassulacean acid metabolism (CAM) and has been used for thousands of years in medicine, cosmetics, and as an ornamental plant such as was described in the Bible. *Aloe vera* stomata are open at night and closed during the day. The CO₂ acquired by *Aloe vera* at night is temporarily stored as malic and other organic acids and is decarboxylated the following day to provide CO₂ for fixation in the Calvin-Benson cycle behind closed stomata. *Aloe vera* is a model for the study of plant electrophysiology with crassulacean acid metabolism.

It is well known that DC and AC electrostimulation of plants can induce activation of ion channels and ion transport, gene expression, activation of enzymatic systems, electrical signaling, plant movements, enhanced wound healing, plant-cell damage, and plant growth (see for a review [26,27]). Recently, we analyzed the anisotropy and nonlinear properties of electrochemical circuits in the leaves of *Aloe vera* [31-33]. Along the conductive bundles, the behavior of *Aloe vera* leaves is highly nonlinear.

Electrostimulation by voltages with an amplitude higher than 2 V applied to the plant causes a drastic change in the leaf in the form of the initial input resistance drop. This change occurs in the conducting bundles and is probably related to the opening of voltage-gated ion channels in the *Aloe vera* leaf [31]. There is a strong electrical anisotropy of the *Aloe vera* leaf. In the direction across the conductive bundle, the behavior of the system is completely passive and linear like in a regular electric circuit with constant resistance. Conductance parallel to vascular bundles is two orders of magnitude higher than in the perpendicular direction.

The existence of electrical signaling in plants has been known for more than two centuries [21-25,28-30]. Direct measurements of plant electrical signaling induced by plasma jets turned out to be more complicated due to large electrical signals from plasma jets transmitted in plants.

Prolonged treatment of *Aloe vera* leaves with the plasma ball does not cause any visible changes in the leaves during plasma treatment or after treatment (Figure 3A). The treatment of *Aloe vera* leaves with a cold atmospheric pressure He-plasma jet induces strong damage to leaves (Figure 3B).

Electrostimulation of electrical networks in plants can induce electrotonic or action potentials propagating along their leaves and stems. Both action and electrotonic potentials play important roles in plant physiology and signal transduction between abiotic or biotic stress sensors and.
Cold Atmospheric Pressure Plasma Jet and Plasma Lamp Interaction with Plants: Electrostimulation, Reactive Oxygen and Nitrogen Species, and Side Effects

plant responses. It is well known that electrostimulation of plants can induce activation of ion channels, ion transport, plant-cell damage, enhanced wound healing, gene expression, enzymatic systems activation, electrical signaling, plant movements, and influence plant growth. Electrostimulation is an important tool for the evaluation of mechanisms of phytoactuators' responses in plants without stimulation of abiotic or biotic stress phytosensors.

The plasma ball and CAPPJ caused strong electrical signaling along the leaves. The amplitude of the electric signal induced by the plasma lamp decreased with the distance in the leaf, but the frequency of 22 kHz was constant (Figure 4A). The amplitude of the electrical signal in the leaves of *Aloe vera* could reach several volts (Figure 4A). The propagation of the electric wave along the *Aloe vera* leaf can be illustrated by the equivalent electric diagram in Figure 4B. The mathematical model and experimental analysis of the electrotonic potential transduction in *Aloe vera* were presented earlier by Volkov and Shtessel [33].

Electrical signals can propagate along the plasma membrane over long and short distances in vascular bundles, plasmodesmata, and protoxylems. Electrotonic potentials in plants were discovered recently [31-33]. Electrostimulation of electrical circuits in *Aloe vera* induced electrotonic potentials with amplitude exponentially decreasing along a leaf or a stem.

The propagation of passive electrical signals in plant conductive bundles is usually interpreted in terms of the cable model [34]. The cable theory of the flow of electricity in a leaky cable was created by Lord Kelvin, who derived the equations to study the transatlantic telegraph cable. Hodgkin and Rushton [35] and Rall [36] applied the cable theory to passive electrical flow along membranes. Basic assumptions underlying the cable theory are as follows: (a) the electrotonic potential is due to the change in the membrane potential, which propagates in a cylinder with constant radius; (b) passive electrotonic current is ohmic in accord with the Ohm law; (c) electrotonic current divides between internal and membrane resistance; (d) membrane current is inversely proportional to membrane surface area; (e) axial current is inversely proportional to diameter.

If voltage-gated ion channels are closed and not involved in signal transduction along the plasma membrane, the transduction of electrotonic potentials can be described by the cable theory along a circuit consisting of capacitors C1 and resistors R1 and R2 (Figure 4B).

Plasma ball induced high-frequency electrical signaling in a cabbage leaf with an amplitude of several volts (Figure 5). The frequency of the electrical wave corresponds to the frequency of the electromagnetic field in the plasma ball.
Electrical signals in tomato plants induced by the plasma ball are shown in Figure 6. These results of electrical waves in tomato plants (Figure 6) are very similar to the electrical waves in cabbage (Figure 5) and Aloe vera leaves (Figure 4).

**Cold atmospheric pressure plasma He-jet interaction with plants**

Direct treatment of plants with CAPPJ can damage their tissue (Figure 3B). It can be the effect of RONS, UV-Vis light, and electromagnetic fields produced by the plasma. A thin quartz plate was inserted between the plasma jet and a leaf of Aloe vera in control experiments. UV-Vis light and electromagnetic fields can penetrate through the quartz plate without visibly damaging the leaf (Figure 7). This means that RONS was responsible for damaging the plant tissue in Aloe vera (Figure 3B).

The treatment of cabbage leaves with a cold He-plasma jet caused visible damage around the place of contact of the plasma with the leaf (Figure 8A). The plasma ball treatment does not cause visible damage to the cabbage leaf (Figure 8B,C).

The FTIR and optical diffusive reflectance spectra (Figure 9) show the difference between untreated and treated cabbage leaves when the treatment was done with the He-plasma jet for 10 min. The FTIR spectra have an absorption maximum of 1 (Figure 9A) at about 3200 cm⁻¹ which can be attributed to the phenol O-H stretching groups [37]; peak 2 is from the resonance groups of the C-H aliphatic (between 2850 and 3000 cm⁻¹) [37]; peak 3 at 1603 cm⁻¹ is from the resonance groups of the aromatic C=C [37]; and peak 4 at 1246 cm⁻¹ refers to the C-O stretching from hemicellulose and lignin [37].

Reflectance spectra of Bonnie hybrid cabbage leaf before and after 10 min treatment with cold atmospheric pressure He-plasma jet have a very significant difference between 300 nm and 550 nm (Figure 9B). This is most likely caused by the degradation of chlorophyll by reactive oxygen species in the brown spots of leaves treated with the cold He-plasma jet (Figure 8A).
Discussion

Most publications and patents about the effect of cold plasma on plants focus on a significant increase in crop yield and plant sustainability [2,3,37]. The new terms “plasma seeds” and “plasma agriculture” have been widely used in the last 25 years. Cold plasma can protect surfaces of a bio-tissue against bacteria, viruses, fungi, and mold [38-40]. Here we found for the first time that cold atmospheric pressure He-plasma jet (CAPP) can also cause side effects and damage to plants. UV-Vis radiation, high frequency strong electromagnetic field, RONS, ions, and free electrons from plasma can also generate side effects (Figures 3-8) and changes in the composition of plants and fruits [14,41,42]. Reactive oxygen species can induce plant cell death [43-45]. Ozone can induce necrosis and increase peroxidase activity [46]. The development of side effects depends on the duration of plasma treatment. Plasma disinfects many bacteria in half a minute [39], so the optimal time for processing seeds and plants with cold atmospheric pressure He-plasma jet is in the range of 10 to 60 seconds, although this time also depends on the composition of the gas phase used to produce plasma. We found that the electrostimulation of plants by plasma lamps can help to avoid side processes in biological tissues associated with the generation of RONS.

Conclusion

This article gives a new insight into the possible side effects of cold plasma interactions with plants. Scheme 1 shows mechanisms of interaction of cold atmospheric pressure He-plasma jet and/or plasma lamps with seeds and plants. It is known that CAPP in the air produces RONS at room temperature. Reactive oxygen and nitrogen species play important roles in plant physiology and agriculture. They can be very toxic to biological tissue and can selectively kill bacteria, fungi, and viruses. At the same time, RONS are useful companions of plant developmental processes and the activation of ion channels. RONS are involved in signal transduction, interactions with ion channels, and cell death. The specific biological response of a plant to RONS depends on the chemical identity of the RONS, the intensity of the signal, sites of production, the plant developmental stage, and interactions with other signaling molecules such as nitric oxides, hydrogen peroxide, ozone, and nitric acid. Cold plasma can affect ion transport and bioelectrochemical characteristics of plant tissue. Generated by the cold atmospheric pressure He-plasma jet reactive oxygen and nitrogen species, UV-Vis photons, and high-frequency strong electromagnetic fields with amplitudes of a few kV can interact with plants. Here we found that RONS produced by CAPPJ can also cause side effects and damage to plants if the plasma exposure is long enough. The plasma lamp has no visible side effects on Aloe vera, cabbage, and tomato plants, but induces electrical waves with very high amplitude in plants. The plasma ball creates high-frequency electromagnetic fields that can be used for electroporation and corrugation of biological tissues. Understanding the mechanisms of plasma interactions with seeds and plants can contribute to the development of plasma-based technology to control plant developmental, increase yield and growth rates, and protect from pathogens. Low-temperature atmospheric pressure plasma can play an important role in agriculture, medicine, food processing, disinfection and sterilization, and biophysical and biochemical applications.

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Author contributions

A.G.V. conceived the idea, analyzed the data, and participated in manuscript writing. All authors provided the data and edited the manuscript. All authors have read and agreed to the published version of the manuscript.

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Cold Atmospheric Pressure Plasma Jet and Plasma Lamp Interaction with Plants: Electrostimulation, Reactive Oxygen and Nitrogen Species, and Side Effects


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Cold Atmospheric Pressure Plasma Jet and Plasma Lamp Interaction with Plants: Electrostimulation, Reactive Oxygen and Nitrogen Species, and Side Effects


